

# Robert A. Cross

Centre for Mechanochemical Cell Biology

Warwick Medical School

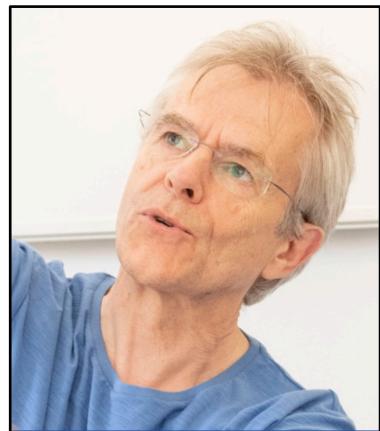
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Born 16.04.57, Duffield, Derbyshire, UK



## EDUCATION

1983 PhD in Protein Biophysics (University of Nottingham)

1979 BSc in Biochemistry (University of Nottingham)

## POSITIONS

2018 - 2022 Director, MSci Integrated Natural Sciences programme, Warwick

2009 - Director, Centre for Mechanochemical Cell Biology, Warwick

2009 - Professor of Mechanochemical Cell Biology, Warwick Medical School

1991 - 2009 Leader, Molecular Motors Group, Marie Curie Research Institute, UK

1988 - 1991 Research Staff, MRC LMB, Cambridge, UK

## FELLOWSHIPS

1986 - 1988 MDA Fellow MRC LMB, Hills Rd, Cambridge CB2 2QH

1984 - 1986 EMBO Fellow OAW IMB, 5020 Salzburg, Austria

## CURRENT SUPPORT

2020 - 2025 Wellcome Trust £1.66M Investigator award  
*Interlocking nanomechanics of kinesins and tubulins*

## AWARDS

2024 Biochemical Society award for Sustained Excellence

## COMMITTEE WORK

2015-2018 Wellcome ERG8 Molecular Basis of Cell Function

## SCIENTIFIC INTERESTS

My lab studies the molecular mechanisms by which kinesins and microtubules generate force. I aim to contribute new understanding of this core aspect of eukaryotic self-organisation. Recently, I have been interested in how the mechanical cycles of kinesins and tubulins are coupled to one another, and how allosteric effectors affect this coupling. The view that kinesins and microtubules sense and adapt to each other's mechanical cycles is a paradigm shift for which we are partly responsible. My goal for the foreseeable future is to define how kinesins and tubulins detect, respond to and manipulate each other's mechanics and dynamics. Many drugs that are important for human health and for agriculture target the machinery of microtubule-based transport. Our work builds understanding of the mechanisms by which these drugs exert their effects on the transport machinery.

My lab also engineers new types of microscope and we continue to develop the Warwick Open Source Microscope, a hardware platform for advanced optical microscopy, aiming to benefit the global open microscopy community ([www.wosmic.org](http://www.wosmic.org)).

## BIOSKETCH

I am a molecular motors enthusiast. I obtained my PhD in **1983** from Nottingham University and then won an EMBO long-term fellowship to work with J. Victor Small and Apolinary Sobieszek in Salzburg on smooth muscle myosin. I made seminal discoveries regarding myosin self assembly (see below). In **1986** I moved to MRC-LMB as an MDA fellow and, working with John Kendrick Jones, Clive Bagshaw and Mike Geeves, was able to formulate an explicit mechanism for myosin II self-assembly. In **1991** I moved to Marie Curie Cancer Research Institute and began working on kinesin, then newly-discovered. We established key features of the stepping mechanisms of several kinesins. In **2009**, the MCRI closed and I moved, with Andrew McAinsh and Anne Straube and most of our lab members, to Warwick University, to co-found a new centre for mechanochemical cell biology, a term we coined. My current work reflects a paradigm shift in which we recognise that kinesins and microtubules sense and adapt to each other's mechanochemical cycles. We have discovered that kinesins make backslips rather than backsteps, and that kinesin strong binding stabilises and expands the lattice of GDP-microtubules. Most recently, we found that taxol has different effects on different human tubulin isotypes, and that ATP $\gamma$ S drives well-coordinated kinesin stepping under load.

## TRAINING RECORD

Every person who has worked with me since I started my own lab in 1991 has been supported in their further development. A full list is at <https://mechanochemistry.org/Cross/alumni.php>. PDRAs and students who have left my lab have been supported in taking away their projects and materials and I have further supported them closely in establishing themselves as independent scientists. All the PhD students I have supervised have graduated successfully. Examples:

**Kurt Anderson** (PDRA, 1998-2000) was my postdoc and now leads the centre for advanced light microscopy at the Crick Institute.

**Andrew Lockhart** (PDRA, 1991-1996) was my first postdoc. He is now head of GSK R&D Neuroscience China.

**Junichiro Yajima** (PDRA, 2003-2008) and I developed several novel single molecule methodologies for analysis of the kinesin mechanism. Jun is now an Associate Professor with his own lab in Tokyo University.

**Miho Katsuki** (PDRA, 2006-2012) worked with me on microtubule polymerases and microtubule lattice conformation. Miho now has her own lab in Fukuoka University.

**Douglas Drummond** (PDRA/sPDRA, 2003-2015) was my right-hand man for many years. Doug and Miho Katsuki married in 2012 and Doug moved to a full professorship at Kyushu University in Fukuoka.

**Algirdas Toleikis** (PDRA 2015-2020) discovered that kinesins slip backwards rather than stepping backwards. Algis won a highly prestigious EMBO Installation Grant to start his own lab in Lithuania.

I have trained 7 technicians. **Christina Hoey** (RA 2002-2005), **Michael Osei** (RA 2005-2007), **Virginia (Wai Chun) Tse** (RA 2019-2020) and **Rebecca Sipthorpe** (RA 2019-2020) all went on to take up PhD studentships. **Thilani Babuji** (RA 2021-2023) just started her PhD at Bristol. **Naomi Shepherd** (RA 2018-2021) left to focus on powerlifting and is now world champion in her weight category.

I am in touch with everybody. **Jun Yajima**, **Doug Drummond** and **Algiris Toleikis** are current collaborators.

## CONFERENCE ORGANISATION

I have organised a number of international conferences on molecular motors:

- |           |   |
|-----------|---|
| 2018      | <b>CMCB-Randall</b> meeting “ <i>Mechanobiology at the Shard II</i> ”               |
| 2017      | <b>CMCB-Randall</b> meeting “ <i>Mechanobiology at the Shard</i> ”                  |
| 2013      | <b>BSCB</b> Spring meeting “ <i>Mechanochemical Cell Biology</i> ”                  |
| 2012      | <b>JSPS</b> Japan-UK international symposium on <i>Mechanochemical Cell Biology</i> |
| 2008      | <b>MCRI</b> Spring Workshop Microtubule Dynamics                                    |
| 2005      | <b>EMBO</b> workshop / Harden Conference in Cambridge, UK <i>Molecular Motors</i>   |
| 1993-2008 | <b>Marie Curie</b> international workshops on <i>Molecular Motors</i>               |
| 1998      | <b>FEBS</b> Advanced Course at MCRI, <i>Molecular Motors</i>                        |
| 1993      | <b>EMBO</b> workshop in Cambridge, UK <i>Molecular Motors</i>                       |
| 1991      | <b>Wellcome Trust</b> Frontiers in Science congress <i>Molecular Motors</i>         |
| 1990      | <b>EMBO</b> workshop in Maria Alm, <i>Smooth Muscle Contraction</i>                 |

## OTHER CONTRIBUTIONS TO RESEARCH CULTURE

The Centre for Mechnochemical Cell Biology at Warwick ([www.mechanochemistry.org](http://www.mechanochemistry.org)), of which I am founding director, hosts currently 20 research groups from 5 Warwick departments (Life Sciences, Physics, Maths, Computing, Medical School) all of whom interrogate the mechanochemical mechanisms of living systems. My goal has been to develop an optimal physical and intellectual research environment for this work and we are succeeding in this.

The CMCB was an early signatory of SF-DORA and we continue to promote Open Science. I am much invested also in educating the next generation of mechanochemical cell biologists, via postgraduate training and also, from **2020**, via an innovative undergraduate course at Warwick, CF10 MSci Integrated Natural Sciences, of which I am co-founder and co-director. This course is intended as a UK version of the Integrated Science programs at Princeton and Harvard, aiming to teach undergraduates practical quantitative biology *sans frontiers*.

In connection with this course, my lab has developed an educational version of our Warwick Open Source Microscope, the eduWOSM. Both researchers and educators are adopting and developing our designs, including internationally.

I am a Wellcome Investigator and until recently was a member of the Wellcome Expert Review Group in the Molecular Basis of Cell Function.

## FUNDING RECORD

2020 - 2025	<b>Wellcome £1.68M Investigator award</b>
2023 – 2024.	<b>BBSRC ALERT 22 £767K</b> – bioAFM with Mishima (PI)
2017 - 2021	<b>Leverhulme Trust £256K</b> – with Anne Straube & Marco Polin
2015 -	<b>BBSRC ALERT14 £0.74M</b> – with Royle (PI), McAinsh/Smith/Cross/Frigerio
2014 - 2019	<b>Wellcome Trust £1.2M Senior Investigator award</b>
2013 - 2017	<b>WPHCT £100K Warwick Open Source Microscope (<a href="http://WOSMic.org">http://WOSMic.org</a>)</b>
2013 - 2017	<b>BBSRC £0.55M Chemical Biology of Microtubules (IPA with Syngenta)</b>
2009-2012	<b>EPSRC £0.2M Programmable kinesin shuttle (with A. Turberfield, Oxford)</b>
2009-2012	<b>BBSRC £0.2M Kinesin-DNA chimeras (with A. Turberfield, Oxford)</b>
2009-2012	<b>AICR £0.2M Single molecule analysis of microtubule tip tracking <i>in vitro</i></b>
2009-	<b>Marie Curie Cancer Care £1.23M transitional programme funding</b>
2004-2009	<b>MRC Strategic Grant £0.5M High resolution structure-function of tubulin</b>
2006-2009	<b>CRUK £0.15M Reconstitution of <i>S. pombe</i> microtubule dynamics <i>in vitro</i></b>
2003-2009	Co-holder on <b>EPSRC Nanotechnology</b> network grant <b>£9M</b>
1999-2003	<b>HFSP</b> network grant ~ <b>£1M</b> (between 5 labs)

Between 1991 and 2009, work in my MCRI group was consistently rated Alpha-A at external quincennial review using the MRC scales (the highest possible category). Marie Curie core funding allowed me to maintain a lab of 4-6 people and to pursue important long-term projects such as the construction of our optical trap.

Whilst at MCRI we were mostly ineligible for external funding. When opportunities opened, I generally applied and mostly succeeded in winning grants. From 1999-2003 we were part of a really excellent Human Frontiers Science Programme network Dynamics of the kinesin tubulin interface involving Hideo Higuchi, Joe Howard, Sharyn Endow, Keiko Hirose and myself. This grant was ranked first out of 250 applications in its round. In 2003 the MRC briefly ran a scheme called Strategic Grants, which allowed them to fund programmes of exceptional merit that fell outside the normal rules – we won a 5 yr strategic grant in a falling market to work on tubulin structure function in collaboration with Linda Amos and Jan Löwe at MRC-LMB. I obtained an SERC project grant when I first set up my lab in 1991, but MCRI was subsequently declared ineligible. In 2002 I tried an application to BBSRC and my proposal was exceptionally well reviewed, but then disallowed. In 2007, I sought to submit a LoLa application to BBSRC, but again MCRI eligibility proved an insurmountable hurdle. In 2005, CRUK agreed to consider one application from MCRI and our project to reconstitute *S. pombe* microtubule dynamics was fully funded, after which we were again shut out. In 2009 AICR made Miho Katsuki and I a 3 year project grant to carry on this work. This grant was amongst the 7% of grants funded in that round. We were invited to join a £9M bionano IRC based in Oxford Physics. Following on from the IRC, Andrew Turberfield and I obtained a responsive-mode EPSRC grant to continue work that used DNA self-assembly to control kinesin-driven motility. A related BBSRC application was also successful. Both these grants were held at Oxford, but the 3 workers split their time between our labs.

In late 2009, I was awarded £1.23M transitional programme funding from Marie Curie Cancer Care, in a competitive process again involving external peer review. This award funded my startup at Warwick. The move to Warwick, completed in Feb 2010, enabled us to create a new centre, the Centre for Mechanochemical Cell Biology, of which I am director. We built a new building to house the centre. UoW contributed £3.4M, AWM (the regional development agency) contributed £2.2M and the Wolfson Trust contributed £1M. Construction began in Sept 2010 and completed one year later. UoW also provided £1.5M towards the purchase of microscopy and other equipment for the new centre. Three years later we extended the building, fully funded by UoW.

## MOST SIGNIFICANT CONTRIBUTIONS

- 2023 YM Chew, RA Cross (2023)  
**Taxol acts differently on different tubulin isotypes**  
Communications Biology 6 (1), 946  
• Reveals that taxol changes the conformation of only some human tubulin isotypes. Our work has the potential to influence practice.
- 2020 A Tolekis, NJ Carter, RA Cross (2020)  
**Backstepping mechanism of kinesin-1**  
Biophysical Journal 119 (10), 1984-1994  
• Shows that kinesin-1 backsteps are not strides, but slips.
- 2018 Peet DR, Burroughs NJ, Cross RA (2018)  
**Kinesin expands and stabilizes the GDP- microtubule lattice**  
Nature Nanotechnology 13 386-391  
• Shows that kinesin is a powerful allosteric effector for tubulin that stabilizes and expands the microtubule lattice.
- 2016 Britto M, Goulet A, Rizvi S, von Loeffelholz O, Moores, CA, Cross RA (2016)  
**S. pombe kinesin-5 switches direction using a steric blocking mechanism**  
Proceedings of the National Academy of Sciences 113, E7483-E7489  
• Shows that physical crowding of kinesin-5 motors on the *S. pombe* microtubule lattice reverses their stepping action.
- 2011 Grant BJ, Gheorghe DM, Zheng W, Alonso M, Huber G, Dlugosz M, McCammon JA , Cross RA (2011)  
**Electrostatically biased binding of kinesin to microtubules**  
PLoS Biology 9 (11), e1001207 d  
• Incipient kinesin motors deviate towards the plus end just before they bind to microtubules.
- 2007 Alonso MC, Drummond DR, Kain S, Hoeng J, Amos LA & Cross RA (2007)  
**An ATP-gate controls tubulin binding by the tethered head of kinesin-1**  
Science 316 120-123  
• ATP binding to one kinesin head regulates the ability of the other to bind to tubulin. This trigger mechanism provides the basis for controlling the stepping action of kinesin.
- 2005 Carter NJ & Cross RA (2005)  
**Mechanics of the kinesin step**  
Nature 435 308-12  
• Using single molecule optical trapping, we discovered that kinesin-1 can step processively backwards under load.
- 2005 Yajima J & Cross RA (2005)  
**A torque component in the kinesin-1 power stroke**  
Nature Chemical Biology 1 338-341  
• Kinesin uses ATP turnover to generate impulses of force and movement. We establish here that this impulse includes an off-axis component.
- 2004 Crevel I, Nyitray M, Weiss S, Geeves MA, Cross RA (2004)  
**What kinesin does at roadblocks: the coordination mechanism for molecular walking**  
EMBO J 23 23-32  
• Establishes that kinesins that hit an obstacle retry forward stepping multiple times before dissociating.
- 2000 Drummond DR & Cross RA (2000)  
**Dynamics of interphase microtubules in *Schizosaccharomyces pombe***  
Current Biology 10 766-775  
• Pioneering quantitative cell biological study, establishing that microtubules catastrophise under compression at cell ends, blazing the trail for much subsequent work.

- 1996 Hirose, K., Lockhart, A. Cross, R.A. and Amos,L.A. (1996)  
**Three dimensional cryoEM of dimeric kinesin and ncd motor domains on microtubules**  
PNAS 93 9539-9544  
• With Linda Amos & Keiko Hirose, pioneering cryoEM reconstructions of kinesin states.
- 1995 Hirose K, Lockhart A., Cross RA & Amos LA (1995)  
**Nucleotide-dependent angular change in kinesin motor domain bound to tubulin**  
Nature 376 277-279  
• With Linda Amos & Keiko Hirose, we did pioneering EM of kinesin states.
- 1994 Lockhart A and Cross RA (1994)  
**Origins of reversed directionality in the ncd molecular motor**  
EMBO J. 13 751-757  
• Establishes that ncd, a reverse-directed kinesin, binds the same microtubule site and executes much the same chemical kinetic cycle as kinesin-1.
- 1992 TP Hodge, R Cross, J Kendrick-Jones (1992)  
**Role of the COOH-terminal nonhelical tailpiece in the assembly of a vertebrate nonmuscle myosin rod**  
The Journal of cell biology 118 (5), 1085-1095  
• Follows up on our 1986 discovery that smooth muscle myosin self-assembly is orchestrated by a C-terminal tailpiece.
- 1988 RA Cross, AP Jackson, S Citi, J Kendrick-Jones, CR Bagshaw (1988)  
**Active site trapping of nucleotide by smooth and non-muscle myosins**  
Journal of molecular biology 203 (1), 173-181  
• Extends our earlier finding with smooth muscle myosin to nonmuscle myosins. Later, we extended further, to skeletal muscle myosins.
- 1986 RA Cross, KE Cross, A Sobieszek (1986)  
**ATP-linked monomer-polymer equilibrium of smooth muscle myosin: the free folded monomer traps ADP. Pi**  
The EMBO journal 5 (10), 2637-2641  
• Reveals that formation of the compact 10s conformation of myosin traps the active site nucleotide, so that assembly is a requirement for ATP turnover.
- 1986 RA Cross, J Vandekerckhove (1986)  
**Solubility-determining domain of smooth muscle myosin rod**  
FEBS letters 200 (2), 355-360  
• I discovered that a C-terminal section of the myosin tail is required for filament assembly

## PUBLICATIONS 1983-2023

Google scholar [https://scholar.google.co.uk/citations?hl=en&user\\_= iaVZaQAAAAJ](https://scholar.google.co.uk/citations?hl=en&user_= iaVZaQAAAAJ)

Chew YM, Cross RA (2023)

**Taxol acts differently on different tubulin isotypes**

Commun Biol. 2023 Sep 16;6(1):946.

doi: 10.1038/s42003-023-05306-y. PMID: 37717119; PMCID: PMC10505170.

Sugawa M, Maruyama Y, Yamagishi M, Cross RA, Yajima J. (2022)

**Motor generated torque drives coupled yawing and orbital rotations of kinesin coated gold nanorods**

Commun Biol. 2022 Dec 20;5(1):1368. doi: 10.1038/s42003-022-04304-w. PubMed PMID: 36539506; PubMed Central PMCID: PMC9767927.

Siddiqui N, Roth D, Toleikis A, Zwetsloot AJ, Cross RA, Straube A. (2022)

**Force generation of KIF1C is impaired by pathogenic mutations**

Curr Biol. 2022 Sep 12;32(17):3862-3870.e6. doi: 10.1016/j.cub.2022.07.029. Epub 2022 Aug 11. PubMed PMID: 35961316; PubMed Central PMCID: PMC9631238.

Toleikis A, Carter NJ, Cross RA (2020)

**Backstepping Mechanism of Kinesin-1**

Biophys J. 2020 Nov 17;119(10):1984-1994. doi: 10.1016/j.bpj.2020.09.034. Epub 2020 Oct 6.

PubMed PMID: 33091340; PubMed Central PMCID: PMC7732724.

Chew YM, Cross RA (2020)

**Molecular Motors: Kif14's Disordered Dongle**

Curr Biol. 2020 Sep 7;30(17):R988-R991. doi: 10.1016/j.cub.2020.06.095. PubMed PMID: 32898497.

von Loeffelholz O, Peña A, Drummond DR, Cross R, Moores CA (2019)

**Cryo-EM Structure (4.5-Å) of Yeast Kinesin-5-Microtubule Complex Reveals a Distinct Binding Footprint and Mechanism of Drug Resistance**

J Mol Biol. 2019 Feb 15;431(4):864-872. doi: 10.1016/j.jmb.2019.01.011. Epub 2019 Jan 16. PubMed PMID: 30659798; PubMed Central PMCID: PMC6378684.

Cross RA (2019)

**Microtubule lattice plasticity**

Curr Opin Cell Biol. 2019 Feb;56:88-93. doi: 10.1016/j.ceb.2018.10.004. Epub 2018 Nov 8. Review. PubMed PMID: 30415187.

Meadows JC, Messin LJ, Kamnev A, Lancaster TC, Balasubramanian MK, Cross RA, Millar JB (2018)

**Opposing kinesin complexes queue at plus tips to ensure microtubule catastrophe at cell ends**

EMBO Rep. 2018 Nov;19(11). doi: 10.15252/embr.201846196. Epub 2018 Sep 11. PubMed PMID: 30206188; PubMed Central PMCID: PMC6216294.

McHugh T, Drechsler H, McAinsh AD, Carter NJ, Cross RA (2018)

**Kif15 functions as an active mechanical ratchet**

Mol Biol Cell. 2018 Jul 15;29(13):1743-1752. doi: 10.1091/mbc.E18-03-0151. Epub 2018 May 17. PubMed PMID: 29771628; PubMed Central PMCID: PMC6080711.

Peet DR, Burroughs NJ, Cross RA (2018)

**Kinesin expands and stabilizes the GDP-microtubule lattice**

Nat Nanotechnol. 2018 May;13(5):386-391. doi: 10.1038/s41565-018-0084-4. Epub 2018 Mar 12. PubMed PMID: 29531331; PubMed Central PMCID: PMC5937683.

von Loeffelholz O, Venables NA, Drummond DR, Katsuki M, Cross R, Moores CA (2017)

**Nucleotide- and Mal3-dependent changes in fission yeast microtubules suggest a structural plasticity view of dynamics**

Nat Commun. 2017 Dec 13;8(1):2110. doi: 10.1038/s41467-017-02241-5. PubMed PMID: 29235477; PubMed Central PMCID: PMC5727398.

Cross R, Veigel C (2017)

**Molecular machines**

Biophys Rev. 2017 Aug;9(4):287-288. doi: 10.1007/s12551-017-0285-3. Epub 2017 Aug 7. PubMed PMID: 28786029; PubMed Central PMCID: PMC5578928.

Britto M, Goulet A, Rizvi S, von Loeffelholz O, Moores CA, Cross RA (2016)

**Schizosaccharomyces pombe kinesin-5 switches direction using a steric blocking mechanism**  
Proc Natl Acad Sci U S A. 2016 Nov 22;113(47):E7483-E7489. doi: 10.1073/pnas.1611581113. Epub 2016 Nov 9. PubMed PMID: 27834216; PubMed Central PMCID: PMC5127321.

Huang J, Chew TG, Gu Y, Palani S, Kamnev A, Martin DS, Carter NJ, Cross RA, Oliferenko S, Balasubramanian MK (2016)

**Curvature-induced expulsion of actomyosin bundles during cytokinetic ring contraction**  
Elife. 2016 Oct 13;5. doi: 10.7554/elife.21383. PubMed PMID: 27734801; PubMed Central PMCID: PMC5077295.

Cross RA (2016)

**Review: Mechanochemistry of the kinesin-1 ATPase**

Biopolymers. 2016 Aug;105(8):476-82. doi: 10.1002/bip.22862. Review. PubMed PMID: 27120111; PubMed Central PMCID: PMC4924600.

Hussmann F, Drummond DR, Peet DR, Martin DS, Cross RA (2016)

**Alp7/TACC-Alp14/TOG generates long-lived, fast-growing MTs by an unconventional mechanism**

Sci Rep. 2016 Feb 11;6:20653. doi: 10.1038/srep20653. PubMed PMID: 26864000; PubMed Central PMCID: PMC4749977.

Andrecka J, Ortega Arroyo J, Lewis K, Cross RA, Kukura P (2016)

**Label-free Imaging of Microtubules with Sub-nm Precision Using Interferometric Scattering Microscopy**

Biophys J. 2016 Jan 5;110(1):214-7. doi: 10.1016/j.bpj.2015.10.055. PubMed PMID: 26745424; PubMed Central PMCID: PMC4806212.

Cross RA, McAinsh A (2014)

**Prime movers: the mechanochemistry of mitotic kinesins**

Nat Rev Mol Cell Biol. 2014 Apr;15(4):257-71. doi: 10.1038/nrm3768. Review. PubMed PMID: 24651543.

Klejnot M, Falnikar A, Ulaganathan V, Cross RA, Baas PW, Kozielski F (2014)

**The crystal structure and biochemical characterization of Kif15: a bifunctional molecular motor involved in bipolar spindle formation and neuronal development**

Acta Crystallogr D Biol Crystallogr. 2014 Jan;70(Pt 1):123-33. doi: 10.1107/S1399004713028721. Epub 2013 Dec 24. PubMed PMID: 24419385; PubMed Central PMCID: PMC3919264.

Katsuki M, Drummond DR, Cross RA (2014)

**Ectopic A-lattice seams destabilize microtubules**

Nat Commun. 2014;5:3094. doi: 10.1038/ncomms4094. PubMed PMID: 24463734; PubMed Central PMCID: PMC3921467.

Wollman AJ, Sanchez-Cano C, Carstairs HM, Cross RA, Turberfield AJ (2014)

**Transport and self-organization across different length scales powered by motor proteins and programmed by DNA**

Nat Nanotechnol. 2014 Jan;9(1):44-7. doi: 10.1038/nnano.2013.230. Epub 2013 Nov 10. PubMed PMID: 24213281; PubMed Central PMCID: PMC3883648.

Carter NJ, Cross RA (2012)

**Kinesin backsteps**

Biochem Soc Trans. 2012 Apr;40(2):400-3. doi: 10.1042/BST20120008. Review. PubMed PMID: 22435819.

Erent M, Drummond DR, Cross RA (2012)

**S. pombe kinesins-8 promote both nucleation and catastrophe of microtubules**

PLoS One. 2012;7(2):e30738. doi: 10.1371/journal.pone.0030738. Epub 2012 Feb 20. PubMed PMID: 22363481; PubMed Central PMCID: PMC3282699.

Kaseda K, McAinsh AD, Cross RA (2012)

**Dual pathway spindle assembly increases both the speed and the fidelity of mitosis**

Biol Open. 2012 Jan 15;1(1):12-8. doi: 10.1242/bio.2011012. Epub 2011 Oct 24. PubMed PMID: 23213363; PubMed Central PMCID: PMC3507165.

Cross RA, McAinsh AD, Straube A (2011)

**Mechanochemical cell biology**

Semin Cell Dev Biol. 2011 Dec;22(9):913-5. doi: 10.1016/j.semcdb.2011.10.002. Epub 2011 Oct 7. PubMed PMID: 22001112.

Grant BJ, Gheorghe DM, Zheng W, Alonso M, Huber G, Dlugosz M, McCammon JA, Cross RA (2011)

**Electrostatically biased binding of kinesin to microtubules**

PLoS Biol. 2011 Nov;9(11):e1001207. doi: 10.1371/journal.pbio.1001207. Epub 2011 Nov 29. PubMed PMID: 22140358; PubMed Central PMCID: PMC3226556.

Katsuki M, Muto E, Cross RA (2011)

**Preparation of dual-color polarity-marked fluorescent microtubule seeds**

Methods Mol Biol. 2011;777:117-26. doi: 10.1007/978-1-61779-252-6\_9. PubMed PMID: 21773925.

Drummond DR, Kain S, Newcombe A, Hoey C, Katsuki M, Cross RA (2011)

**Purification of tubulin from the fission yeast Schizosaccharomyces pombe**

Methods Mol Biol. 2011;777:29-55. doi: 10.1007/978-1-61779-252-6\_3. PubMed PMID: 21773919.

Cross RA (2010)

**Kinesin-14: the roots of reversal**

BMC Biol. 2010 Aug 16;8:107. doi: 10.1186/1741-7007-8-107. PubMed PMID: 20731883; PubMed Central PMCID: PMC2922096.

Katsuki M, Drummond DR, Osei M, Cross RA (2009)

**Mal3 masks catastrophe events in Schizosaccharomyces pombe microtubules by inhibiting shrinkage and promoting rescue**

J Biol Chem. 2009 Oct 23;284(43):29246-50. doi: 10.1074/jbc.C109.052159. Epub 2009 Sep 9. PubMed PMID: 19740752; PubMed Central PMCID: PMC2764634.

Kaseda K, McAinsh AD, Cross RA (2009)

**Walking, hopping, diffusing and braking modes of kinesin-5**

Biochem Soc Trans. 2009 Oct;37(Pt 5):1045-9. doi: 10.1042/BST0371045. PubMed PMID: 19754449.

Braun M, Drummond DR, Cross RA, McAinsh AD (2009)

**The kinesin-14 Klp2 organizes microtubules into parallel bundles by an ATP-dependent sorting mechanism**

Nat Cell Biol. 2009 Jun;11(6):724-30. doi: 10.1038/ncb1878. Epub 2009 May 10. PubMed PMID: 19430466.

des Georges A, Katsuki M, Drummond DR, Osei M, Cross RA, Amos LA (2008)

**Mal3, the Schizosaccharomyces pombe homolog of EB1, changes the microtubule lattice**

Nat Struct Mol Biol. 2008 Oct;15(10):1102-8. doi: 10.1038/nsmb.1482. Epub 2008 Sep 14. PubMed PMID: 18794845; PubMed Central PMCID: PMC2575238.

Cross R (2008)

**Single molecule for the people**

Nat Cell Biol. 2008 Sep;10(9):1014. doi: 10.1038/ncb0908-1014. PubMed PMID: 18758487.

Kaseda K, Crevel I, Hirose K, Cross RA (2008)

**Single-headed mode of kinesin-5**

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